6.2 Sampling

Storage temperature of soil sample, 4°C , is too low to keep plant parasitic nematodes alive, while $10 \cdot 15^{\circ}\text{C}$ is more suitable (Barker and Nusbaum, 1971). This is possibly true for free living nematodes, because the nematodes include some species (e.g., Aphelenchoididae, Anguinidae) taxonomically very close to plant parasitic ones. The fact also supports $10 \cdot 15^{\circ}\text{C}$ as the storage temperature, that *C. elegans* researchers usually choose this temperature range rather than 4°C , to maintain the worms for long.

6.3 Extraction

Oostenbrink elutriator is expensive and difficult to set up in some laboratories. In addition, the equipment is commercially produced only by one company in the world, which may cause unstable supply. Such equipment should not be designated in an international standard procedure.

Bearmann funnel/tray extraction without prior soil elutriation, is still a popular method worldwide (Forge and Simard, 2001; Fu, et al., 2000; Hanel, 2001; Hoschitz and Kaufmann, 2004; Okada, et al., 2004; Ruess, et al., 2001; Sohlenius, 2002; Yeates, et al., 1999). Usually free living nematodes do not have cysts or inactive stage, so that Bearmann funnel/tray extraction is effective to obtain the nematodes, although the extraction efficiency may differ between nematode taxa, as is often the case with any type of extraction method (McSorley and Frederick, 2004). One of the advantages of Bearmann extraction is that the nematode suspension obtained includes few or no debris which will hinder nematode identification seriously. More importantly, the method is simple, inexpensive and less laborious to operate, which makes it available worldwide. Such a method is suitable as an international standard. Soil elutriation prior to the extraction should not be designated obligatory, but optional.

Cotton-wool filters are not available in some countries. Cotton-linter filters, or paper wipers such as Kim Wiper (Kimberly-Clark) are more available world wide.

6.4 Counting

In counting nematodes, the general procedure is acceptable, but the size of bottle (100 ml) to keep nematode suspension, and the volume of suspension for counting (10 ml) should be optional. This is because almost all nematodes from the sample soil should be counted if the nematode density is too low, while only 1% of suspension may includes too many nematode individuals if the original density in soil is very high. Thus, degree of dilution or concentration of the original suspension, and % of suspension to be

examined should be changed depending on the nematode density.

Nematodes just fixed in formalin may lack key characteristics for generic level identification. Thus, formalin fixation should be just minimum requirement. Transfer of nematodes into anhydrous glycerol by Seinhorst's rapid method (Seinhorst, 1959) should be recommended for more promising identification.

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